





First Synthesis and Pharmacological Evaluation of Benzoindolizidine and Benzoquinolizidine Analogues of α - and β -Peltatin

Axel Couture,^{a,*} Eric Deniau,^a Pierre Grandclaudon,^a Stéphane Lebrun,^a Stéphane Léonce,^b Pierre Renard^c and Bruno Pfeiffer^c

^aLaboratoire de Chimie Organique Physique, ESA CNRS 8009, Université des Sciences et Technologies de Lille, F-59655 Villeneuve d'Ascq Cedex, France

bInstitut de Recherches Servier, Division de Cancérologie Expérimentale, 11 rue des Moulineaux, F-92415 Suresnes, France c'ADIR, 1 rue Carle Hébert, F-92415 Courbevoie Cedex, France

Received 20 March 2000; accepted 19 May 2000

Abstract—The benzoindolizidine and-quinolizidine analogues of α - and β -peltatin were designed and synthesized by two different synthetic routes involving as the key step the Bischler–Napieralski cyclization of suitably substituted *N*-acyl-2-arylmethylpyrrolidine and -piperidine derivatives. The in vitro biological activity of these analogues as well as some of their derivatives was subsequently evaluated. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Podophyllotoxin 1 and related compounds, including α and β-peltatin 2 and 3 (Fig. 1) are a class of cyclolignan type compounds characterized by several vicinal oxygen functions on the differently substituted aromatic moieties and/or in the central carbocyclic unit. They have been obtained from the rhizoma resin of *Podophyllum peltatum* L^2 and have long been known to display anti-tumour³ and mitotoxic activities.^{4,5} However due to toxic sideeffects they have in most cases failed to give satisfactory clinical trial results. 6–9 In fact, the clinical utility of these compounds is compromised by the highly reactive nature of the fused γ -lactone moiety present in these molecules and by epimerization at C2 under physiological conditions. 10 This has been shown to be deleterious to the therapeutic action by giving rise to products devoid of anti-tumour activity. 11 Consequently, a number of groups have carried out modifications at this stereogenic centre and new analogues which are incapable of loss of configurational integrity at the C2 centre, which encompass a broad spectrum of antineoplastic activities and thus overcome clinical limitations, have been explored. 12-17 In particular, ingenious variations have been proposed

by several groups which have notably designed and synthesized a variety of analogues in which the sp3 carbon is replaced by a sp2 nitrogen, anticipating configurational integrity and avoiding any problems of epimerization. ^{18–24} Paradoxically, despite the fact that they retain anti-tumour activities, to our knowledge, such modifications have been mainly confined to the aza-analogues of podophyllotoxin $\mathbf{4-6}^{1,18-26}$ (Fig. 1) and the study of the α and β -peltatin congeners appears to be an unexplored area. We wish therefore to report in this paper the first synthesis and the in vitro biological evaluation of hexahydropyrrolo[1,2-b] and hexahydropyrido[1,2-b]isoquinoline analogues of α and β -peltatin 7–10.

Results and Discussion

Chemistry

Two complementary and undoubtedly transposable methods were adopted for the synthesis of the benzoin-dolizidine and -quinolizidine analogues of α and β -peltatin 7, 8 and 9, 10 respectively (Fig. 1). The first one which is depicted in the retrosynthetic Scheme 1 involved as the key step the formation of the enamides 13, 14 which could be derived from the Horner reaction between the appropriate *O*-benzyl protected aromatic carboxal-dehyde 15 and the phosphorylated *N*-acyl pyrrolidine

^{*}Corresponding author. Tel.: +33-(0)3-20-43-44-32; fax: +33-(0)3-20-33-63-09; e-mail: axel.couture@univ-lille1.fr

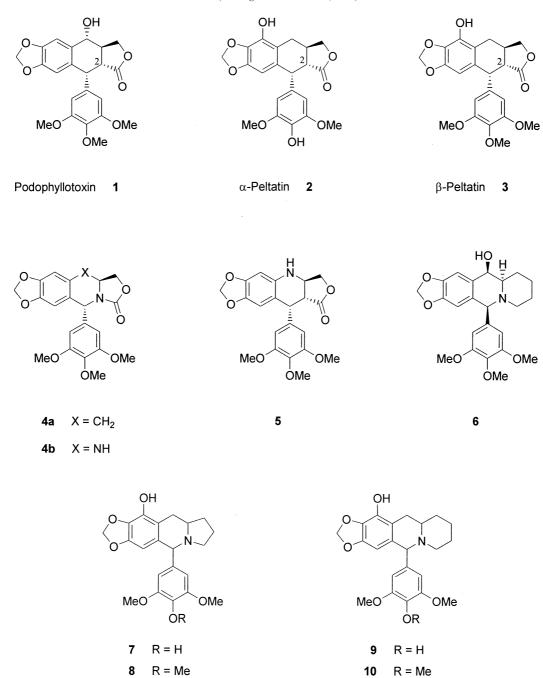


Figure 1.

derivatives **16**, **17** respectively. Reduction and subsequent Bischler–Napieralski cyclization then should complete the synthesis of the target compounds **7**, **8**.

It was obvious that this synthetic strategy would be fraught with difficulties associated with the presence of several hydroxy phenolic functions at different sites on the environmentally different aromatic moieties of the final compounds 7, 8. Simple retrosynthetic analysis indicates that the hydroxy phenolic function at the western part of the molecule should be incorporated via the trialkoxybenzaldehyde derivative 15 whereas the corresponding function on the pendant aromatic unit of 7 should be integrated into the models via the protected 1-aroyl-2-diphenylphosphinoylpyrrolidine derivative 16.

The synthesis started with the preparation of the benzyl protected 4-hydroxybenzo[d][1,3]dioxole-5-carbaldehyde **15**. To reach this protected phenol derivative, the metallation and subsequent hydroxylation of piperonal was investigated. Critical to the success of this strategy was the ability to identify a masked carbonyl synthon that was capable not only of retaining the formyl functionality but also of directing subsequent lithiation at the 'in between' ring position. The metallation conditions described by Comins for various aldehydes by using lithium N,N,N'-trimethylethylenediamine²⁷ were tested on piperonal but failed. After considerable experimentation we found that adding 2 equiv of t-butyllithium in pentane to 1 equiv of imidazolidine 18^{28} in Et₂O at -78 °C and then quenching with B(OMe)₃ at 0 °C followed by

hydrogen peroxide led to the target compound after regeneration of the formyl functionality and ultimate benzyl protection (Scheme 2). The second partners, 16 and 17, involved in the Horner reaction were easily obtained by coupling the phosphorylated amine 20 with the aromatic carboxylic acids 22, 23, the former being obtained

by sequential protection-oxidation of syringaldehyde (Scheme 3). Initially the phosphorylated pyrrolidine **20** was readily obtained by addition of diphenylphosphine oxide **26** to the triazine **24**.^{29,30} This protocol delivered the phosphorylated carboxamides **16**, **17** in 84 and 78% yield respectively. Horner reaction between compounds **16**, **17**

$$OH OH OH OH OH OH$$

$$OH OH OH OH OH$$

$$OH OH OH OH OH$$

$$OH OH OH OH OH$$

$$MeO OR$$

$$T R = H$$

$$R = H$$

$$R = Me$$

$$11 R = H$$

$$R = Me$$

$$12 R = Me$$

Scheme 1.

n = 1

n = 2

n = 1 (94%)

n = 2 (97%)

DCC , DMAP ,
$$\mathrm{Et_3N}$$
 , $\mathrm{CH_2Cl_2}$

Phi P Ph N

R = Bn (84%)

R = Me (78%)

Scheme 3.

R = Bn

R = Me

(*E*) + (*Z*) R = Bn (71%)

14 (E) + (Z) R = Me (69%)

R = H (87%)

R = Me (90%)

and 15 was then carried out by smooth deprotonation at -78 °C with *n*-BuLi in THF and subsequent treatment with the suitably substituted aldehyde 15. Warming the reaction mixture to room temperature ensured the completion of the reaction and the *N*-acylenamides 13, 14 were isolated in high yields (71% and 69% respectively) as a mixture of *Z*- and *E*-isomers with *E*-isomers predominating (40:60 for 13, 45:55 for 14) (Scheme 4). The stereochemistry was unambiguously assigned from the ¹H NMR spectra and further confirmed by a one-dimensional difference nuclear Overhauser experiment. Thus, in contrast to the *E*-isomer, irradiation of the vinylic proton of the *Z*-form led to an increased intensity of the allylic protons of the pyrrolidine ring.

We then focused on the hydrogenation of the polysubstituted enamides **13** and **14**. The rarely employed method making use of Pd on C and ammonium formate^{31,32} could offer, if feasible, a double advantage by effecting simultaneously the reduction of the enamine function with concomitant removal of the benzyloxy protecting group thus giving straightforward access to direct candidates for the Bischler–Napieralski reaction. As anticipated, treatment of enamides **13**, **14** delivered the reduced di- and monophenolic compounds 11 and 12 with excellent yields (87 and 90% respectively) (Scheme 4). Compounds 11, 12 were then subjected to Bischler-Napieralski conditions by standard treatment with phosphorus oxichloride (POCl₃) in toluene at reflux and subsequent reduction with sodium borohydride (NaBH₄) in methanol. Unfortunately all attempts to obtain the cyclocondensed products by this protocol, even under mild conditions, 23,33 were unrewarding probably due to the presence of the hydroxy phenolic groups in the opened compounds, a precedented complicating factor in related systems.¹⁷ Assuming that a reprotection step might be a prerequisite to the carboannulation reaction to give the target molecules 7 and 8, compounds 11 and 12 were then fully benzyl protected prior to effecting the Bischler-Napieralski reaction (Scheme 5). Gratifyingly, this protocol afforded the expected cyclized products 29 and 30, in 73 and 70% yield respectively, as a mixture of diastereomers in the ratio 10:1. The structure of the major diastereoisomer was deduced from the ¹H NMR spectrum, namely by the presence of long range coupling between the mono and dibenzylic protons attributable to their axial relationship. 34,35 Final removal of the benzyl protecting group of the major isomers delivered

the benzoindolizidine analogues of α - and β -peltatin 7 and 8 with very acceptable yields (46 and 45% respectively over the last three steps).

For the synthesis of the benzoquinolizidine analogues of α - and β -peltatin 9 and 10 (Fig. 1) we decided to adopt an alternative strategy which hinges upon the same synthetic principles but precludes the rather unsatisfactory and tedious deprotection-reprotection steps. This alternative approach which is depicted in the Scheme 6 involves as the key step the formation of the 2-arylmethylpiperidine derivative 33 readily obtained after deprotection and subsequent reduction of enecarbamate 32. This N-protected enamine 32 was derived from the Horner reaction between the aromatic carboxaldehyde 15 and the phosphorylated cyclic carbamate 31. Initially compound 31 was prepared by treatment of the corresponding phosphorylated cyclic amine 21 (Scheme 2) with di-tert-butyldicarbonate.^{29,30} Horner reaction proceeded uneventfully to afford the *N-tert*-butoxycarbonyl-2-arylmethylpiperidine **32** as a

mixture of Z- and E-isomer (5:95). N-Deprotection of the enecarbamate 32 with trifluoroacetic acid and subsequent reduction of the transient iminium ion was performed as a single, one-pot reaction and this procedure afforded straightforwardly the 2-arylmethylpiperidine 33 in fairly good yield. Schotten—Baumann reaction between amine 33 and the appropriate carboxylic acid chlorides 34, 35 furnished the amides 36, 37 candidates for the Bischler–Napieralski reaction. The O-benzyl protections in 36, 37 survived the cyclization conditions since treatment with POCl₃ followed by NaBH₄ delivered the cyclocondensed products 38 and 39 as a mixture of diastereoisomers (90:10 for 38 and 95:5 for 39). Final removal of the benzyl protection in 38, 39 completed the synthesis of the target products 9, 10.

Pharmacological evaluation

Eleven analogues of α and β -peltatin and particularly their urethane derivatives (Scheme 7) were prepared in

Scheme 7.

both the hexahydropyrrolo[1,2-b] and in the hexahydropyrido[1,2-b]isoquinoline series and evaluated in vitro on L1210 leukemia cell line. The results expressed as IC₅₀ (concentration reducing by 50% the cell proliferation) are reported in Table 1. The perturbation of the cell cycle induced by the most active compounds were also investigated on the same cell line (Figure 2 and Table 2).

Surprisingly, the strict analogue of α -peltatin is more active in the pyrido[1,2-b]isoquinoline series (IC₅₀ (L1210)=2.3 μ M for compound **9** versus 5.4 μ M for compound **7**) whilst the strict analogue of β -peltatin is 15 times more potent in the pyrrolo[1,2-b]isoquinoline series (IC₅₀ (L1210)=0.6 μ M for compound **8** versus 9.5 μ M for compound **10**).

Without any exception, all the mono or di O-benzylated intermediates were found inactive or significantly less active than their deprotected analogues (IC₅₀ between 9.7 and 19.3 μ M for compounds **29**, **30**, **38**, **39**).

Table 1. Antiproliferative activity of the target compounds 7–10 and some of their derivatives

Compound	$IC_{50} (\mu M)^a$
Podophyllotoxin 1	0.025
7	5.4
8	0.6
9	2.3
10	9.5
29	9.7
30	17.3
38	18.4
39	19.3
40	4.1
41	0.86
42	9.9

^aInhibition of L1210 cell proliferation measured by the MTT assay (mean of at least 2 values obtained in separate experiments).

By contrast, introduction of a *para*-fluorophenylcarbamate group on the β -peltatin analogues has no influence in both the pyrido[1,2-b]isoquinoline (IC₅₀ (L1210) = 9.9 μ M versus 9.5 μ M for compound **42** and **10**) and the pyrrolo[1,2-b]isoquinoline (IC₅₀ (L1210) = 0.86 μ M versus 0.6 μ M for compound **41** and **8**) series. This is the same thing for the disubstituted analogue of α -peltatin **40**.

Due to their good cytotoxicity, compounds 8 and 41 were selected for investigation of their interaction with the cell cycle of the L1210 cells.

Compared to untreated control, both induced a partial accumulation of the cells on the G_2+M and $>G_2+M$ (probably 8N) phases of the cell cycle at $2.5\,\mu M$ as reported on Figure 2 and Table 2. Between 65% and 69% of the cells are in the G_2M phase of the cell cycle compared to 24% for untreated control. About 20% are in the 8N phase (tetraploid cells) compared to 1% for untreated control. The DNA histogram of compounds 8 and 41 is comparable to that obtained with podophyllotoxin at $0.025\,\mu M$. Accumulation of the cell in the tetraploid state could be compatible with antimitotic properties.

Table 2. Typical DNA histogram and distribution into the different phases of the cell cycle of untreated (light) or treated (heavy) L1210 cells

Compound	% of L1210 cells in cell cycle phases	
	$G_2 + M (\mu M)$	$> G_2 + M (8N) (\mu M)$
Untreated control Podophyllotoxin 8 41	24% 55% (0.025) 65% (2.5) 69% (2.5)	1% 26% (0.025) 21% (2.5) 20% (2.5)

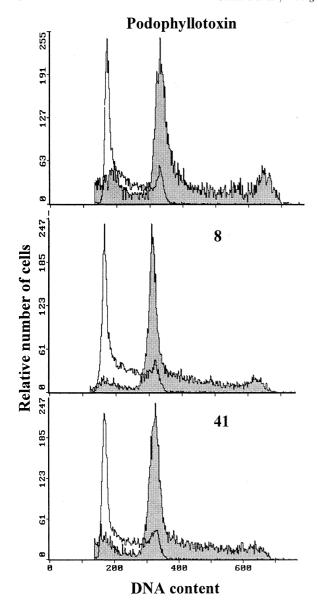


Figure 2. Typical DNA histogram and distribution into the different phases of the cell cycle of untreated (light) or treated (heavy) L1210 cells.

Experimental

General information

Methanol was distilled from magnesium turnings. Tetrahydrofuran (THF) and ether (Et₂O) were pre-dried with anhydrous Na₂SO₄ and distilled over sodium benzophenone ketyl under Ar before use. CH₂Cl₂, NEt₃, toluene were distilled from CaH₂. Dry glassware was obtained by oven-drying and assembly under dry Ar. The glassware was equipped with rubber septa and reagent transfer was performed by syringe techniques. For flash chromatography, Merck silica gel 60 (230-400 mesh ASTM) was used. The melting points were taken on a Reichert-Thermopan apparatus and are not corrected. Unless otherwise stated, proton, carbon and phosphorus NMR spectra were taken with CDCl₃ as solvent at 300, 75 and 121 MHz respectively on a Bruker AM 300 spectrometer. Microanalyses were performed by the CNRS microanalysis centre.

Starting materials

Triazines 24,³³ 25²⁹ and diphenylphosphine oxide 26³⁶ were prepared according to the literature methods. The phosphorylated amines 20,³⁰ 21,²⁹ amides 16, 17²⁹ and carbamate 31^{29,30} were synthesized according to already reported procedures. The benzyl protected aromatic carboxylic acid 22³⁷ was obtained by oxidation with Jones reagent of the corresponding aldehyde³⁸ deriving from syringaldehyde. The carboxylic acid chloride 34³⁹ was easily obtained by treatment of the corresponding acid 22 with thionyl chloride and was used directly without further purification.

Phosphorylated amide 16. Prepared in 84% yield as described;²⁹ mp 153–154 °C; IR (KBr) ν: 1646 (CO) and 1161 cm⁻¹ (PO); ¹H NMR: δ 1.79–1.94 (m, 1H, CH₂), 1.96–2.17 (m, 1H, CH₂), 2.18–2.33 (m, 1H, CH₂), 2.47–2.63 (m, 1H, CH₂), 3.33–3.45 (m, 2H, NCH₂), 3.72 (s, 6H, OCH₃), 4.95 (s, 2H, OCH₂Ph), 5.43–5.52 (m, 1H, CH-P), 6.40 (s, 2H, H_{arom}), 7.21–7.43 (m, 7H, H_{arom}), 7.48–7.58 (m, 4H, H_{arom}), 7.79–7.91 (m, 2H, H_{arom}), 8.04–8.16 (m, 2H, H_{arom}); ¹³C NMR: δ C 170.0 (CO), 153.1, 137.4, 132.2, 131.8, 131.1, 130.5 (d, $J_{\rm CP}$ =92.5 Hz), CH 131.4 (d, $J_{\rm CP}$ =9 Hz), 128.8 (d, $J_{\rm CP}$ =11.5 Hz), 128.7, 128.4, 128.1, 128.0 (d, $J_{\rm CP}$ =8 Hz), 104.6, 56.1 (d, $J_{\rm CP}$ =71.5 Hz), CH₂ 74.9, 50.7, 25.6, 24.9, CH₃ 56.1; ³¹P NMR: δ 34.6. Anal. calcd for C₃₂H₃₂NO₅P: C, 70.97; H, 5.96; N, 2.59. Found: C, 71.12; H, 5.88; N, 2.63.

Phosphorylated amide 17. Prepared in 78% yield as described;²⁹ mp 148–149 °C; IR (KBr) v: 1638 (CO) and 1168 cm⁻¹ (PO); ¹H NMR: δ 1.82–1.94 (m, 1H, CH₂), 1.97–2.15 (m, 1H, CH₂), 2.19–2.30 (m, 1H, CH₂), 2.45–2.60 (m, 1H, CH₂), 3.31–3.59 (m, 2H, NCH₂), 3.73 (s, 3H, OCH₃), 3.76 (s, 3H, OCH₃), 3.82 (s, 3H, OCH₃), 5.53–5.56 (m, 1H, CH-P), 6.28 (s, 2H, H_{arom}), 7.31–7.45 (m, 3H, H_{arom}), 7.47–7.55 (m, 3H, H_{arom}), 7.82–7.90 (m, 2H, H_{arom}), 8.05–8.11 (m, 2H, H_{arom}); ¹³C NMR: δ C 170.1 (CO), 152.8, 146.3, 131.5 (d, J_{CP} = 94 Hz), 130.6, CH 131.8 (d, J_{CP} = 8 Hz), 131.4 (d, J_{CP} = 9 Hz), 128.9 (d, J_{CP} = 9 Hz), 128.2 (d, J_{CP} = 10 Hz), 104.6, 56.0 (d, J_{CP} = 79 Hz), CH₂ 50.6, 25.5, 24.9, CH₃ 60.8, 55.9; ³¹P NMR: δ 34.6. Anal. calcd for C₂₆H₂₈NO₅P: C, 67.09; H, 6.02; N, 3.01. Found: C, 67.18; H, 5.93; N, 2.88.

Synthesis of the benzaldehyde derivative 15. The synthesis of benzaldehyde 15 was carried out as follows. A solution of piperonal (5 g, 33.3 mmol) and N,N'-dimethylethylenediamine (3.22 g, 36.6 mmol) in toluene (100 mL) was refluxed for 5 h. The resulting mixture was washed with H_2O . The organic layer was dried over MgSO₄ and the solvent was removed in vacuo to leave an oil which was purified by distillation under reduced pressure to afford the imidazolidine 18 (5.8 g, 79%, bp $108 \,^{\circ}$ C/ $5 \, 10^{-3} \, \text{torr}$).

Imidazolidine 18. ¹H NMR: δ 2.15 (s, 6H, NCH₃), 2.46–2.53 (m, 2H, NCH₂), 3.14 (s, 1H, NCH), 3.32–3.38 (m, 2H, NCH₂), 5.92 (s, 2H, OCH₂O), 6.73 (d, J=7.8 Hz, 1H, H_{arom}), 6.81 (dd, J=7.8, 1.6 Hz, 1H, H_{arom}), 6.99 (d, J=1.6 Hz, 1H, H_{arom}); ¹³C NMR: δ C 147.9, 147.8, 133.8, CH 122.5, 108.4, 107.5, 92.1, CH₂ 100.9, 53.1,

CH₃ 39.4. Anal. calcd for $C_{12}H_{16}N_2O_2$: C, 65.43; H, 7.32; N, 12.72. Found: C, 65.56; H, 7.31; N, 12.61.

To a stirred solution of imidazolidine 18 (2 g, 9.1 mmol) in Et₂O (50 mL) was added t-BuLi (10.7 mL, 1.7 M in pentane, 18.2 mmol) with stirring under Ar at -78 °C. The solution was stirred at -78 °C for 15 min, then warmed to rt over a period of 30 min and re-cooled to -78 °C. A solution of trimethyl borate (0.62 g, 10.9 mmol) was added dropwise by syringe and the mixture was warmed to 0°C over a period of 1 h. Glacial AcOH (0.5 mL) was added followed by hydrogen peroxide (30%, 1.2 mL) and the reaction mixture was stirred at room temperature for 16 h. The crude product was then poured onto an aqueous 6N HCl solution (50 mL) and the mixture was stirred for an additional 30 min. The aqueous layer was extracted twice with CH₂Cl₂ (2×30 mL) and the organic extract subsequently washed with aqueous NaOH (10%, 2×40 mL). Acidification with dilute HCl was followed by extraction with CH_2Cl_2 (2×50 mL). The organic layer was then washed with water and brine and dried (MgSO₄). The dried extract was concentrated in vacuo to afford **19** (1.08 g, 70%) as a yellow solid, mp 115-116 °C (lit.: 40 115–116 °C). Benzylation of 19 under standard procedure furnished quantitatively the benzyl protected benzaldehyde derivative 15.41

Synthesis of the N-acyl enamines 13, 14. A solution of n-BuLi (1.6 M in hexanes, 1.7 mL, 2.75 mmol) was added dropwise to a solution of the phosphorylated amide 16, 17 (2.5 mmol) in THF (30 mL) at -78 °C with stirring under Ar. The orange solution was stirred for an additional 15 min and a solution of the appropriate aldehyde 15 (2.5 mmol) in THF (5 mL) was then added slowly. After being stirred at -78 °C for 15 min the reaction mixture was allowed to come to room temperature over 2h. Aqueous NH₄Cl was added and the organic layer was separated, rinsed with brine, dried (MgSO₄) and concentrated to dryness. The crude products were analyzed by ¹H NMR in order to determine the E/Z-isomers ratio and compounds 13, 14 (E- and Z-isomers) were separated by flash column chromatography using AcOEt: hexane (60:40) as eluent and finally recrystallized from hexane-toluene.

N-Acyl enamine 13. (71%); (*E*)-Isomer, mp 125-126 °C; ¹H NMR: δ 1.78–1.91 (m, 2H, CH₂), 2.73 (td, J=7.6, 1.9 Hz, 2H, = CCH₂), 3.70 (t, J = 7.0 Hz, 2H, NCH₂), 3.79 (s, 6H, OCH₃), 5.02 (s, 2H, OCH₂Ph), 5.17 (s, 2H, OCH_2Ph), 5.92 (s, 2H, OCH_2O), 6.52 (d, J=8.1 Hz, 1H, H_{arom}), 6.67 (d, J=8.1 Hz, 1H, H_{arom}), 6.77 (s, 2H, H_{arom}), 7.25–7.51 (m, 11H, $10H_{arom} + H_{vinyl}$); ¹³C NMR: δ C 169.1 (CO), 153.4, 147.7, 139.9, 139.7, 138.1, 137.5, 137.2, 133.0, 130.6, 124.9, CH 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 121.8, 108.9, 104.6, 102.1, CH₂ 101.0, 75.0, 73.6, 50.4, 30.1, 22.4, CH₃ 56.2; (Z)-Isomer, mp 117– 118 °C; ¹H NMR: δ 1.92–2.20 (m, 2H, CH₂), 2.63 (t, $J = 7.4 \,\mathrm{Hz}, 2H, = \mathrm{CCH}_2$, 3.61 (s, 6H, OCH₃), 3.78 (t, J = 7.1 Hz, 2H, NCH₂), 4.94 (s, 2H, OCH₂Ph), 4.98 (s, 2H, OCH₂Ph), 5.69 (s, 1H, H_{vinyl}), 5.78 (s, 2H, OCH₂O), 6.35 (s, 2H, H_{arom}), 6.38 (d, J = 8.1 Hz, 1H, H_{arom}), 6.57 (d, J = 8.1 Hz, 1H, H_{arom}), 7.28–7.49 (m, 10H, H_{arom}); ¹³C NMR: δ C 168.6 (CO), 152.4, 147.7, 139.1, 138.6, 138.1, 137.5, 136.1, 130.9, 130.5, 123.3, CH 128.6, 128.4, 128.3, 128.2 127.8, 127.7, 123.3, 108.0, 104.5, 102.7, CH₂ 100.7, 74.9, 73.0, 48.6, 32.2, 20.7, CH₃ 55.6. Anal. calcd for $C_{35}H_{33}NO_7$: C, 72.54; H, 5.70; N, 2.42. Found: C, 72.36; H, 5.58; N, 2.63.

N-Acyl enamine 14. (69%); (*E*)-Isomer, mp 116–117 °C; ¹H NMR: δ 1.79–1.88 (m, 2H, CH₂), 2.72 (td, J=7.3, $2.2 \text{ Hz}, 2H, = \text{CCH}_2$, $3.69 \text{ (t, } J = 6.8 \text{ Hz}, 2H, \text{ NCH}_2$), 3.82(s, 6H, OCH₃), 3.84 (s, 3H, OCH₃), 5.18 (s, 2H, OCH₂Ph), 5.92 (s, 2H, OCH₂O), 6.49 (d, J = 8.1 Hz, 1H, H_{arom}), 6.65(d, J = 8.1 Hz, 1H, H_{arom}), 6.74 (s, 2H, H_{arom}), 7.32–7.46 (m, 6H, 5H_{arom}+H_{vinyl}); ¹³C NMR: δ C 169.8 (CO), 153.1, 147.7, 139.9, 139.8, 137.2, 133.6, 130.5, 128.6, 124.8, CH 128.3, 128.0, 127.9, 121.8, 108.9, 104.5, 102.7, CH₂ 101.0, 73.6, 50.9, 30.2, 22.4, CH₃ 60.9, 56.2; (Z)-isomer, mp 96–97 °C; ¹H NMR: δ 1.92–2.13 (m, 2H, CH₂), 2.64 (t, $J = 7.0 \,\mathrm{Hz}, 2 \,\mathrm{H}, = \mathrm{CCH}_2$, 3.63 (s, 6H, OCH₃), 3.82 (s, 3H, OCH_3), 3.84 (t, J=7.3 Hz, 2H, NCH_2), 4.95 (s, 2H, OCH₂Ph), 5.67 (s, 1H, H_{vinyl}), 5.83 (s, 2H, OCH₂O), 6.33 (s, 2H, H_{arom}), 6.37 (d, $J = 8.0 \,\text{Hz}$, 1H, H_{arom}), 6.53 (d, $J = 8.0 \,\text{Hz}$, 1H, H_{arom}), 7.32–7.41 (m, 5H, H_{arom}); ¹³C NMR: δ C 169.8 (CO), 152.8, 147.8, 145.4, 141.8, 136.1, 133.7, 131.0, 130.1, 129.5, CH 127.5, 121.1, 112.9, 108.0, 104.7, 102.6, CH₂ 100.6, 73.7, 49.8, 31.9, 21.1, CH₃ 60.8, 56.3. Anal. calcd for C₂₉H₂₉NO₇: C, 69.17; H, 5.80; N, 2.78. Found: C, 68.91; H, 5.89; N, 2.60.

Synthesis of the *N*-acyl pyrrolidine derivatives 11, 12. A suspension of compounds 13, 14 (2 mmol) in methanol (30 mL) was stirred with activated Pd/C (10%, 20 mg) and a solution of HCOONH₄ (640 mg, 10 mmol) in distilled water (5 mL) was slowly added. The reaction mixture was refluxed for 2 h, filtered on Celite[®] and water was added. Extraction with CH₂Cl₂ (3×20 mL), drying over MgSO₄ and concentration in vacuo left an oily product which was purified by flash chromatography using AcOEt:hexane (1:1) as eluent.

N-Acyl pyrrolidine 11. (87%); mp 168–169 °C; ¹H NMR: 81.75–1.87 (m, 2H, CH₂), 1.93–2.17 (m, 2H, CH₂), 2.36 (dd, J=13.9, 9.6 Hz, 1H, CH₂Ar), 3.44 (d, J=13.9 Hz, 1H, CH₂Ar), 3.55–3.66 (m, 2H, NCH₂), 3.87 (s, 6H, OCH₃), 3.96–4.05 (m, 1H, NCH), 5.93 (s, 2H, OCH₂O), 6.31 (d, J=7.8 Hz, 1H, H_{arom}), 6.48 (d, J=7.8 Hz, 1H, H_{arom}), 6.84 (s, 2H, H_{arom}); ¹³C NMR: 8C 170.4 (CO), 148.0, 146.7, 140.8, 137.1, 134.9, 126.5, 99.8, CH 121.9, 120.0, 105.1, 59.5, CH₂ 101.2, 51.0, 36.8, 31.3, 25.0, CH₃ 56.5. Anal. calcd for C₂₇H₂₃NO₇: C, 62.84; H, 5.73; N, 3.49. Found: C, 62.92; H, 5.79; N, 3.39.

N-Acyl pyrrolidine 12. (90%); mp 185–186 °C; ¹H NMR: δ 1.68–1.79 (m, 2H, CH₂), 1.83–2.01 (m, 2H, CH₂), 2.33 (dd, J=14.1, 9.5 Hz, 1H, CH₂Ar), 3.35 (d, J=14.1 Hz, 1H, CH₂Ar), 3.42–3.51 (m, 2H, NCH₂), 3.79 (s, 6H, OCH₃), 3.85 (s, 3H, OCH₃), 3.75–3.83 (m, 1H, NCH), 5.89 (s, 2H, OCH₂O), 6.25 (d, J=8.0 Hz, 1H, H_{arom}), 6.41 (d, J=8.0 Hz, 1H, H_{arom}), 6.80 (s, 2H, H_{arom}); ¹³C NMR: δ C 170.0 (CO), 147.8, 146.2, 140.4, 137.9, 134.2, 127.1, 100.0, CH 122.3, 121.6, 104.9, 58.9, CH₂ 100.9, 50.1, 37.5, 29.6, 23.2, CH₃ 56.8. Anal. calcd for C₂₂H₂₅NO₇: C, 63.61; H, 6.07; N, 3.37. Found: C, 63.50; H, 6.18; N, 3.31.

Synthesis of the benzylated compounds 27, 28. Compounds 27, 28 were prepared by benzylation of the parent phenolic compounds 11, 12 under standard conditions (BnBr, KOH, MeOH).

Compound 27. (74%); oil; ¹H NMR: δ (mixture of two rotational isomers A and B, 70:30) 1.51-1.98 (m, 4H, CH_2), 2.31–2.46 (m, 3H, $CH_2Ar + 1H NCH_2$, B), 2.59– 2.66 (m, 1H, NCH₂, B), 2.88 (dd, J = 13.2, 7.9 Hz, 1H, CH_2Ar , A), 3.15 (dd, J = 13.2, 3.7 Hz, 1H, CH_2Ar , A), 3.26–3.38 (m, 2H, NCH₂, A), 3.67 (s, 6H, OCH₃, B), 3.74 (s, 6H, OCH₃, A), 4.32–4.39 (m, 1H, NCH, B), 4.46-4.57 (m, 1H, NCH, A), 5.01 (dd, J=15.9, 10.8 Hz, 4H, OCH₂Ph, A), 5.26 (dd, J = 14.9, 11.3 Hz, 4H, OCH₂Ph, B), 5.86 (s, 2H, OCH₂O, B), 5.92 (s, 2H, OCH_2O , A), 6.14 (d, J = 7.7 Hz, 1H, H_{arom} , B), 6.36 (d, J = 7.7 Hz, 1H, H_{arom}, B), 6.41–6.53 (m, 2H, H_{arom}, A), 6.73 (s, 2H, H_{arom}, A), 6.75 (s, 2H, H_{arom}, B), 7.18–7.49 (m, 10H, H_{arom}); ¹³C NMR: δ (mixture of two rotational isomers A and B, 70:30) C 169.6 (CO), 153.2, 148.2, 147.7, 141.6, 137.6, 137.4, 132.9, 132.8, 129.4, CH 128.9, 128.1, 128.0, 127.9, 127.4, 124.5, 123.9, 123.2, 104.8, 104.2, 102.9, 102.6, 58.3 (A), 57.1 (B), CH₂ 100.9 (A), 100.2 (B), 75.0, 73.6, 50.4 (A), 45.6 (B), 34.9 (B), 32.1 (A), 29.3 (B), 29.2 (A), 24.8 (A), 22.7 (B), CH₃ 56.2 (A), 55.2 (B). Anal. calcd for C₃₅H₃₅NO₇: C, 71.79; H, 5.98; N, 2.39. Found: C, 71.66; H, 6.09; N, 2.51.

Compound 28. (80%); mp 61–62 °C; ¹H NMR: δ (mixture of two rotational isomers A and B, 77:23) 1.46-2.05 (m, 4H, CH₂), 2.28-2.43 (m, 3H, CH₂Ar + 1H NCH₂),B), 2.52–2.64 (m, 1H, NCH₂, B), 2.84 (dd, J=13.2, 8.0 Hz, 1H, CH₂Ar, A), 3.09 (dd, J = 13.2, 3.4 Hz, 1H, CH₂Ar, A), 3.17–3.35 (m, 2H, NCH₂, A), 3.56–3.61 (m, 1H, NCH, B), 3.71 (s, 3H, OCH₃), 3.78 (s, 6H, OCH₃), 4.34-4.52 (m, 1H, NCH, A), 4.81 (dd, J=11.9 Hz, 2H, OCH_2Ph , A), 5.58 (dd, J = 15.9, 11.6 Hz, 2H, OCH_2Ph , B), 5.78 (s, 2H, OCH₂O, B), 5.81 (s, 2H, OCH₂O, A), 6.13 (d, $J = 7.6 \,\text{Hz}$, 1H, H_{arom} , B), 6.29 (d, $J = 7.6 \,\text{Hz}$, 1H, H_{arom}, B), 6.33–6.47 (m, 2H, H_{arom}, A), 6.66 (s, 2H, H_{arom}, B), 6.68 (s, 2H, H_{arom}, A), 7.13–7.45 (m, 5H, H_{arom}); ¹³C NMR: δ (mixture of two rotational isomers A and B, 77:23) C 168.9 (CO), 152.4, 147.5, 140.6, 136.9, 136.7, 134.8, 132.4, 132.3, CH 128.4, 127.9, 127.3, 123.8 (A), 123.4 (B), 107.7 (A), 107.6 (B), 102.9 (A), 102.2 (B), 59.1 (B), 58.2 (A), CH₂ 101.6 (B), 100.9 (A), 73.6 (A), 72.6 (B), 50.4 (A), 45.2 (B), 35.3 (B), 32.1 (A), 29.7 (B), 29.1 (A), 24.8 (A), 22.1 (B), CH₃ 60.8, 56.2. Anal. calcd for C₂₉H₃₁NO₇: C, 68.91; H, 6.14; N, 2.77. Found: C, 68.78; H, 6.18; N, 2.90.

Synthesis of the cyclocondensed products 29, 30. A mixture of compound 27, 28 (2 mmol), POCl₃ (1.53 g, 10 mmol) in dry toluene (30 mL) was refluxed for 6 h under Ar with stirring. The solvent and excess reagent were removed under vacuum and the residue was dissolved in dry methanol (20 mL). Sodium borohydride (0.38 g, 10 mmol) was then added portionwise until pH 9, AcOEt (50 mL) was added and the organic layer washed with aqueous NaOH (10%) and dried (MgSO₄). Evaporation of the solvent left an oily residue which was analyzed by ¹H NMR spectroscopy. Major isomer was finally purified by flash column chromatography using

acetone:petroleum ether (1:1) as eluent and by recrystallization from Et₂O:hexane.

Compound 29 (major isomer). Mp 122–123 °C; ¹H NMR: δ 1.59–1.87 (m, 3H, CH₂), 2.03–2.16 (m, 2H, CH₂), 2.34–2.56 (m, 2H, 1H NCH₂+1H CH₂Ar), 2.87 (td, J=9.4, 2.4 Hz, 1H, NCH₂), 3.12 (dd, J=15.9, 2.4 Hz, 1H, NCH), 3.78 (s, 6H, OCH₃), 4.05 (s, 1H, NCHAr), 5.03 (s, 2H, OCH₂Ph), 5.29 (dd, J=16.8, 11.7 Hz, 1H, OCH₂Ph), 5.85 (dd, J=7.6, 1.3 Hz, 2H, OCH₂O), 5.96 (s, 1H, H_{arom}), 6.52 (s, 2H, H_{arom}), 7.27–7.51 (m, 10H, H_{arom}); ¹³C NMR: δ C 153.4, 147.0, 139.7, 139.0, 138.0, 137.7, 136.0, 134.4, 133.5, 121.1, CH 128.5, 128.4, 128.0, 127.9, 127.8, 127.3, 106.2, 102.3, 72.3, 60.8, CH₂ 100.6, 74.9, 73.2, 53.8, 31.2, 30.7, 21.3, CH₃ 56.2. Anal. calcd for C₃₅H₃₅NO₆: C, 74.34; H, 6.19; N, 2.48. Found: C, 74.18; H, 6.03; N, 2.55.

Compound 30 (major isomer). Mp 172–173 °C; 1 H NMR: δ 1.51–1.89 (m, 3H, CH₂), 2.01–2.17 (m, 2H, CH₂), 2.33–2.56 (m, 2H), 2.86 (t, J= 7.9 Hz, 1H), 3.09 (d, J= 14.9 Hz, 1H, NCH), 3.82 (s, 6H, OCH₃), 3.85 (s, 3H, OCH₃), 4.04 (s, 1H, NCHAr), 5.28 (dd, J= 16.8, 11.2 Hz, 2H, OCH₂Ph), 5.84 (d, J= 5.6 Hz, 2H, OCH₂O), 5.94 (s, 1H, H_{arom}), 6.52 (s, 2H, H_{arom}), 7.32–7.48 (m, 5H, H_{arom}); 13 C NMR: δ C 153.1, 147.0, 139.6, 139.0, 137.6, 137.1, 134.4, 133.4, 121.2, CH 128.4, 128.0, 127.8, 106.0, 102.2, 72.2, 60.8, CH₂ 100.7, 73.1, 53.8, 31.2, 30.7, 21.2, CH₃ 60.9, 56.2. Anal. calcd for C₂₉H₃₁NO₆: C, 71.16; H, 6.34; N, 2.86. Found: C, 71.39; H, 6.51; N, 2.73.

Synthesis of the benzoindolizidine analogues of α - and β -peltatin 7, 8. A suspension of compounds 29, 30 (major isomer, 0.5 mmol) and Pd/C (10%, 15 mg) in dry MeOH (20 mL) was treated with a solution of ammonium formate (320 mg, 5 mmol) in distilled water (3 mL). The mixture was refluxed for 2 h, filtered on Celite® and concentrated in vacuo. CH₂Cl₂ (30 mL) and water (30 mL) were added, the organic layer was rinsed with brine and dried (Na₂SO₄). The crude residue was purified by recrystallization from EtOH to afford compounds 7, 8 as pale vellow crystals.

Compound 7. (85%); mp 270–271 °C; ¹H NMR (d_6 -DMSO): δ 1.51–1.85 (m, 3H, CH₂), 2.00–2.21 (m, 2H, CH₂), 2.31–2.75 (m, 3H, NCH₂+1H CH₂Ar), 3.17 (d, J=15.8 Hz, 1H, NCH), 3.77 (s, 6H, OCH₃), 4.04 (s, 1H, NCHAr), 5.88 (s, 2H, OCH₂O), 6.01 (s, 1H, H_{arom}), 6.50 (s, 2H, H_{arom}); ¹³C NMR: δ C 153.3, 146.7, 139.0, 138.2, 135.4, 134.2, 132.8, 99.3, CH 118.1, 105.8, 72.0, 60.3, CH₂ 100.8, 53.4, 30.9, 30.5, 21.0, CH₃ 56.1. Anal. calcd for C₂₁H₂₃NO₆: C, 65.45; H, 5.97; N, 3.64. Found: C, 65.56; H, 5.89; N, 3.80.

Compound 8. (80%); mp 257–258 °C; ¹H NMR (d_6 -DMSO): δ 1.51–1.86 (m, 3H, CH₂), 2.05–2.19 (m, 2H, CH₂), 2.28–2.50 (m, 2H), 2.68–281 (m, 1H), 2.97–3.02 (m, 1H, NCH), 3.81 (s, 6H, OCH₃), 3.84 (s, 3H, OCH₃), 4.07 (s, 1H, NCHAr), 5.68 (s, 1H, H_{arom}), 5.86 (s, 1H, OCH₂O), 6.56 (s, 2H, H_{arom}); ¹³C NMR: δ C 152.7, 145.9, 139.6, 137.2, 136.1, 132.2, 99.0, CH 117.9, 105.7, 70.9, 60.1, CH₂ 100.4, 53.1, 30.8, 30.1, 20.8, CH₃ 60.0, 55.8. Anal. calcd for C₂₂H₂₅NO₆: C, 66.16; H, 6.27; N, 3.51. Found: C, 66.01; H, 6.36; N, 3.62.

Synthesis of the enecarbamate 32. Enecarbamate 32 was prepared by metallation of the N-protected phosphorylated piperidine 31³⁰ and subsequent treatment with aromatic aldehyde 15 as already described for enamides 13, 14 and obtained in 80% yield as a mixture of (E)- and (Z)isomers (95:5). (E)-Isomer; mp 101–102 °C; ¹H NMR (C_6D_6) : δ 1.27–1.38 (m, 4H, CH₂), 1.42 (s, 9H, CH₃), 2.21–2.27 (m, 2H, CH₂), 3.51–3.58 (m, 2H, CH₂), 5.17 (s, 2H, OCH₂Ph), 5.37 (s, 2H, OCH₂O), 6.44 (d, J = 8.0 Hz, 1H, H_{arom}), 6.60 (s, 1H, H_{vinyl}), 6.67 (d, $J = 8.0 \,Hz$, 1H, H_{arom}), 7.02–7.18 (m, 3H, H_{arom}), 7.33–7.37 (m, 2H, H_{arom}); ¹³C NMR (C_6D_6): δ C 154.3, 148.9, 141.0, 139.7, 138.2, 124.1, 79.2, CH 128.6, 128.3, 123.5, 120.5, 103.1, CH₂ 101.1, 73.8, 47.1, 28.3, 26.0, 25.6, CH₃ 28.5. Anal. calcd for C₂₅H₂₉NO₅: C, 70.90; H, 6.90; N, 3.31. Found: C, 70.96; H, 6.78; N, 3.26.

Synthesis of the 2-arylmethylpiperidine derivative 33. Trifluoroacetic acid (1.8 mL, 23 mmol) was added under Ar with stirring to a solution of the enecarbamate **32** (0.973 g, 2.3 mmol) in anhydrous CH₂Cl₂ (5 mL) and stirring was maintained overnight. Evaporation of the solvent and excess reagent under vacuum left a residue which was dissolved in anhydrous MeOH (50 mL). Sodium borohydride (885 mg, 23.3 mmol) was then added portionwise at 0 °C and the reaction mixture was stirred at this temperature for an additional 1 h. EtOAc (60 mL) was then added and the organic solution was washed twice with aqueous NH₄OH (2×50 mL) then with water and brine and finally dried over Na₂SO₄. The solvent was removed under vacuum to leave an oily residue of the compound 33 (0.56 g, 75%) which was used without further purification; ¹H NMR (C_6D_6): δ 1.61– 1.80 (m, 7H, CH₂ + NH), 2.63–2.81 (m, 2H, CH₂), 2.92– 2.98 (m, 1H, CH₂), 3.21 (br. t, J = 6.7 Hz, 1H, CH₂), 3.83 (dd, J=8.9, 3.1 Hz, 1H, CH), 5.23 (s, 2H, OCH₂Ph), 5.89(s, 2H, OCH₂O), 6.46 (d, J = 7.9 Hz, 1H, H_{arom}), 6.62 (d, $J = 7.9 \,\mathrm{Hz}$, 1H, H_{arom}), 7.21–7.48 (m, 5H, H_{arom}); ¹³C NMR (C₆D₆): δ C 147.8, 140.5, 137.5, 128.6, 125.9, CH 128.3, 128.0, 127.8, 123.0, 102.8, 59.5, CH₂ 100.8, 73.4, 45.9, 36.1, 31.1, 25.5, 24.7. Anal. calcd for C₂₀H₂₃NO₃: C, 73.82; H, 7.12; N, 4.30. Found: C, 73.63; H, 6.98; N, 4.44.

Synthesis of the *N*-acyl piperidines 36,37. The Schotten–Baumann reaction between amine 33 and the appropriate carboxylic acid chlorides 34, 35 was carried out under standard conditions.

N-Acyl piperidine 36. (78%); yellow oil; ¹H NMR: δ 1.30–1.96 (m, 6H, CH₂), 2.43–2.57 (m, 2H, CH₂), 2.94 (dd, *J* = 13.5, 8.0 Hz, 1H, CH₂), 3.16–3.31 (m, 2H, CH₂), 3.86 (s, 6H, OCH₃), 5.45 (s, 2H, OCH₂Ph), 5.50 (s, 2H, OCH₂Ph), 5.90 (s, 2H, OCH₂O), 6.21 (s, 1H, H_{arom}), 6.72 (s, 2H, H_{arom}), 7.15–7.43 (m, 10H, H_{arom}); ¹³C NMR: δ C 169.2 (CO), 153.4, 148.1, 147.6, 140.3, 138.0, 137.1, 134.2, 132.1, 131.8, 129.5, CH 129.0, 128.2, 128.1, 128.0, 127.5, 124.5, 107.7, 55.2, CH₂ 100.8, 74.1, 73.2, 45.4, 36.0, 31.1, 25.4, 19.3, CH₃ 56.2. Anal. calcd for C₃₆H₃₇ NO₇: C, 72.59; H, 6.26; N, 2.35. Found: C, 72.71; H, 6.46; N, 2.17.

N-Acyl piperidine 37. (75%); yellow oil; 1 H NMR: δ 1.34–1.98 (m, 6H, CH₂), 2.40–2.56 (m, 2H, CH₂), 2.96 (dd, J = 13.4, 8.1 Hz, 1H, CH₂), 3.12–3.26 (m, 2H, CH₂),

3.79 (s, 6H, OCH₃), 3.82 (s, 3H, OCH₃), 5.51 (s, 2H, OCH₂Ph), 5.91 (s, 2H, OCH₂O), 6.18 (s, 1H, H_{arom}), 6.67 (s, 2H, H_{arom}), 7.12–7.39 (m, 5H, H_{arom}); 13 C NMR: δ C 169.1 (CO), 152.1, 147.8, 140.7, 137.1, 134.5, 132.1, 131.7, CH 128.5, 128.0, 127.5, 123.9, 107.5, 55.1, CH₂ 100.9, 73.1, 45.2, 35.3, 30.9, 25.3, 19.2, CH₃ 60.8, 56.3. Anal. calcd for C₃₀H₃₃NO₇: C, 69.35; H, 6.40; N, 2.70. Found: C, 69.13; H, 6.36; N, 2.64.

Synthesis of the cyclocondensed products 38, 39. The Bischler–Napieralski cyclization reaction of amides 36, 37 was performed as previously described for the corresponding pyrrolidine derivatives 27, 28 to give 38, 39 in 76 and 71% yield respectively.

Compound 38 (major isomer). Mp 149–150 °C; ¹H NMR: δ 1.23–1.88 (m, 7H, CH₂), 2.24 (br. t, J=9.7 Hz, 1H, CH₂), 2.47 (dd, J=16.4, 11.0 Hz, 1H, CH₂), 2.79–2.88 (m, 2H, CH₂), 3.77 (s, 6H, OCH₃), 3.96 (s, 1H, NCH), 5.00 (s, 2H, OCH₂Ph), 5.27 (dd, J=16.0, 11.7 Hz, 2H, OCH₂Ph), 5.82 (dd, J=5.2, 1.2 Hz, 2H, OCH₂O) 5.87 (s, 1H, H_{arom}), 6.50 (s, 2H, H_{arom}), 7.25–7.49 (m, 10H, H_{arom}); ¹³C NMR: δ C 153.4, 146.8, 140.5, 138.3, 138.0, 137.6, 134.0, 132.3, 127.7, 119.9, CH 128.5, 128.4, 128.0, 127.8, 106.4, 102.3, 72.6, 57.8, CH₂ 100.5, 74.9, 73.1, 54.1, 34.0, 31.8, 26.0, 24.4, CH₃ 56.2. Anal. calcd for C₃₆H₃₇NO₆: C, 74.59; H, 6.43; N, 2.42. Found: C, 74.65; H, 6.32; N, 2.30.

Compound 39 (major isomer). Mp 173–174 °C; ¹H NMR: δ 1.26–1.85 (m, 7H, CH₂), 2.26 (t, J=10.5 Hz, 1H, CH₂), 2.45 (dd, J=16.1, 10.5 Hz, 1H, CH₂), 2.75–2.92 (m, 2H, CH₂), 3.82 (s, 6H, OCH₃), 3.84 (s, 3H, OCH₃), 3.96 (s, 1H, NCH), 5.27 (dd, J=16.2, 11.6 Hz, 2H, OCH₂Ph), 5.80 (s, 2H, OCH₂O), 5.91 (s, 1H, H_{arom}), 6.52 (s, 2H, H_{arom}), 7.26–7.44 (m, 5H, H_{arom}); ¹³C NMR: δ C 153.0, 146.8, 140.5, 138.3, 137.6, 136.3, 134.0, 132.2, 119.9, CH 128.4, 128.0, 127.8, 106.2, 102.3, 72.5, 57.7, CH₂ 100.5, 73.1, 54.1, 34.0, 31.7, 26.0, 24.4, CH₃ 60.8, 56.1. Anal. calcd for C₃₀H₃₃NO₆: C, 71.55; H, 6.61; N, 2.78. Found: C, 71.61; H, 6.69; N, 3.73.

Synthesis of the benzoquinolizidine analogues of α - and β -peltatin 9, 10. The final deprotection of the annulated compounds 38, 39 (major isomer) was carried out as described previously for the indolizidine analogues.

Compound 9. (87%); mp 290–291 °C; ¹H NMR: δ 1.50–2.10 (m, 7H, CH₂), 2.85–3.18 (m, 3H, CH₂), 3.46–3.52 (m, 1H, CH₂), 3.75 (s, 6H, OCH₃), 3.83 (s, 1H, NCH), 5.50 (s, 1H, H_{arom}), 5.91 (s, 2H, OCH₂O), 6.95 (s, 2H, H_{arom}); ¹³C NMR: δ C 146.5, 136.7, 136.1, 133.4, 126.8, 124.8, 98.0, CH 115.5, 106.1, 70.8, 60.8, CH₂ 100.9, 52.0, 30.4, 28.2, 22.4, 21.7, CH₃ 56.1. Anal. calcd for C₂₃H₂₇NO₆: C, 66.15; H, 6.31; N, 3.51. Found: C, 66.21; H, 6.36; N, 3.40.

Compound 10. (90%); mp 308–309 °C; ¹H NMR: δ 1.56–2.18 (m, 7H, CH₂), 2.78–3.09 (m, 3H, CH₂), 3.35–3.54 (m, 1H, CH₂), 3.69 (s, 3H, OCH₃), 3.77 (s, 6H, OCH₃), 3.81 (s, 1H, NCH), 5.49 (s, 1H, H_{arom}), 5.92 (s, 2H, OCH₂O), 7.02 (s, 2H, H_{arom}); ¹³C NMR: δ C 146.4, 136.8, 136.1, 133.6, 126.6, 125.0, 98.2, CH 115.6, 106.2,

70.8, 60.8, CH₂ 100.9, 52.0, 34.0, 28.2, 22.4, 21.7, CH₃ 56.1. Anal. calcd for C₂₃H₂₇NO₆: C, 66.81; H, 6.58; N, 3.39. Found: C, 66.74; H, 6.65; N, 3.36.

Synthesis of the urethanes 40–42. Urethanes 40–42 were prepared according to the literature methods.⁴²

Compound 40. (75%); mp 115–116 °C; ¹H NMR: δ 1.51–1.82 (m, 5H, CH₂), 1.96–2.12 (m, 2H, CH₂), 2.33–2.59 (m, 1H, CH₂), 2.75–2.97 (m, 1H, CH), 3.81 (s, 6H, OCH₃), 4.11 (s, 1H, NCH), 5.90 (d, J=5.6 Hz, 2H, OCH₂O), 6.15 (s, 1H, H_{arom}), 6.61 (s, 2H, H_{arom}), 6.96–7.43 (m, 10H, 8H_{arom}+2NH); ¹³C NMR: δ C 156.2 (d, J_{CF}=249 Hz, CF), 154.1 (d, J_{CF}=249 Hz, CF), 152.7, 150.6, 147.1, 142.2, 137.7, 133.9, 133.2, 133.0, 130.5, 122.5, 120.7, 120.5, CH 116.0, 115.8, 115.7, 115.5, 106.0, 105.7, 72.1, 60.3, CH₂ 101.9, 53.7, 31.1, 30.2, 21.2, CH₃ 56.3. Anal. calcd for C₃₅H₃₁F₂N₃O₈: C, 63.73; H, 4.74; N, 6.37. Found: C, 63.87; H, 4.58; N, 6.15.

Compound 41. (70%); mp 157–158 °C; 1 H NMR: δ 1.48–1.79 (m, 5H, CH₂), 1.98–2.11 (m, 2H, CH₂), 2.36–2.65 (m, 1H, CH₂), 2.80–3.03 (m, 1H, CH), 3.82 (s, 6H, OCH₃), 3.85 (s, 3H, OCH₃), 4.06 (s, 1H, NCH), 5.92 (s, 2H, OCH₂O), 6.12 (s, 1H, H_{arom}), 6.54 (s, 2H, H_{arom}), 6.97–7.04 (m, 2H, H_{arom}), 7.27–7.43 (m, 3H, 2H_{arom} + NH); 13 C NMR: δ C 155.9 (d, $J_{\rm CF}$ = 240 Hz, CF), 153.1, 150.5, 147.0, 139.2, 133.4, 130.5, 122.5, 120.7, 120.6, CH 115.9, 115.7, 106.1, 105.9, 72.1, 60.4, CH₂ 101.8, 53.7, 31.2, 30.2, 21.1, CH₃ 60.9, 56.2. Anal. calcd for C₂₉H₂₉FN₂O₇: C, 64.92; H, 5.45; N, 5.22. Found: C, 67.17; H, 5.65; N, 5.06.

Compound 42. (78%); mp 121–122 °C; ¹H NMR: δ 1.28–1.57 (m, 4H, CH₂), 1.71–1.86 (m, 3H, CH₂), 2.30–2.28 (m, 1H, CH₂), 2.51–2.60 (m, 1H, CH₂), 2.67 (dd, J=14.3, 3.5 Hz, 1H), 2.79–2.83 (m, 1H, CH), 3.82 (s, 6H, OCH₃), 3.85 (s, 3H, OCH₃), 3.98 (s, 1H, NCH), 5.86 (dd, J=6.1, 1.4 Hz, 2H, OCH₂O), 6.08 (s, 1H, H_{arom}), 6.55 (s, 2H, H_{arom}), 6.95–7.03 (m, 2H, H_{arom}), 7.36–7.44 (m, 3H, 2H_{arom}+NH); ¹³C NMR: δ C 157.7 (d, J=245 Hz, CF), 153.2, 150.6, 146.8, 140.0, 137.2, 133.3, 132.2, 129.7, 121.3, 120.6, CH 115.9, 115.6, 106.2, 106.0, 72.4, 57.5, CH₂ 101.6, 54.0, 34.0, 31.2, 25.9, 24.3, CH₃ 60.9, 56.2. Anal. calcd for C₃₀H₃₁FN₂O₇: C, 65.45; H, 5.68; N, 5.09. Found: C, 65.71; H, 5.60; N, 5.30.

Biological materials: cell culture and cytotoxicity

L1210 cells (Murine Leukemia) provide by the NCI, Frederik, USA were cultivated in RPMI 1640 medium (Gibco) supplemented with 10% foetal calf serum, 2 mM L-glutamine, 100 units/mL penicillin, 100 μ g/mL streptomycin, and 10mM HEPES buffer (pH = 7.4).

Cytotoxicity was measured by the microculture tetrazolium assay as described. 43 Cells were exposed to graded concentrations of the compounds for 48 h and results expressed as IC₅₀ (concentration which reduced by 50% the optical density of treated cells with respect to untreated controls).

For the cell cycle analysis, L1210 cells (2.5 10⁵ cells/mL) were incubated for 21 h with various concentrations of

the compounds, then fixed by 70% ethanol (v/v), washed and incubated in PBS containing $100 \,\mu g/mL$ RNAse and $25 \,\mu g/mL$ propidium iodide for 30 min at $20 \,^{\circ}$ C. For each sample, 10^4 cells were analyzed on an ATC3000 flow cytometer (Bruker, France) using an argon laser (Spectra-Physics) emitting 400 mW at 488 nM. The fluorescence of propidium iodide was collected through a 615 nM long-pass filter. Data are displayed as linear histograms and results are expressed as the percentage of cells found in the G_2+M phase of the cell cycle.

References

- 1. Ramos, A. C.; Peláez-Lamamié de Clairac, R.; Medarde, M. *Heterocycles* **1999**, *51*, 1443.
- 2. (a) Kotchetkov, N. K.; Kharlin, A. Y.; Ghizhov, O. G.; Sheichenko, V. L. *Tetrahedron Lett.* **1961**, 730. (b) Hartwell, J. L.; Schrecker, A. N. *Fortschr. Chem. Org. Naturst.* **1958**, *15*, 83
- 3. Horwitz, S. B.; Loike, J. D. Lloydia 1977, 40, 82.
- 4. Weiss, S. G.; Tin-Wa, M.; Perdue, R. E.; Farnsworth, N. R. J. Pharm. Sci. 1975, 64, 95.
- 5. Keller-Juslen, C.; Kuhn, M.; von Wartburg, A.; Stehelin, H. *J. Med. Chem.* **1971**, *14*, 936.
- 6. Cortese, F.; Bhattacharyya, B.; Wolff, J. J. Biol. Chem. 1977, 252, 1134.
- 7. King, L. S.; Sullivan, M. S. Science 1946, 104, 244.
- 8. Jardine, I. In *Anticancer Agents Based on Natural Products*; Cassady, J. M.; Douros, J. D., Eds.; Academic: New York, 1980, pp 319–351.
- 9. Ayres, D. C.; Loike, J. D. *Lignans, Chemical, Biological and Clinical Properties*. Cambridge University Press: Cambridge, 1990
- 10. Emmenegger, H.; Stahelin, H.; Rutschmann, J.; Renz, J.; von Wartburg, A. *Arzneim. Forsch* **1961**, *327*, 459.
- 11. Evans, W. E.; Sinkule, J. A.; Horwath, A.; Crom, W. R.; Dow, L. W.; Riviera, G. *Proc. Am. Assoc. Cancer Res.* **1981**, 690
- 12. Ward, R. S. Chem. Soc. Rev. 1982, 11, 75.
- 13. Ward, R. S.. Synthesis, 719.
- 14. Ward, R. S. Nat. Prod. Rep. 1993, 10, 1.
- 15. Ward, R. S. Nat. Prod. Rep. 1995, 12, 183.
- 16. Ward, R. S. Nat. Prod. Rep. 1997, 14, 43.
- 17. Ward, R. S. Nat. Prod. Rep. 1999, 16, 75.
- 18. Tomioka, K.; Kubota, Y.; Koga, K. *Tetrahedron* **1993**, *49*, 1891.
- 19. Lienard, P.; Quirion, J.-C.; Husson, H.-P. *Tetrahedron* **1993**, *49*, 3995.
- 20. Arai, Y.; Enomoto, K. Yakugaku Zasshi 1968, 88, 1197. Chem. Abstr. 1969, 70, 27530.
- 21. Pearce, H. L.; Bach, N. J.; Cramer, T. L. *Tetrahedron Lett.* **1989**, *30*, 907.
- 22. Kadow, J. F.; Vyas, D. M.; Doyle, T. W. Tetrahedron Lett. 1989, 30, 3299.
- 23. van der Eycken, J.; Bosmans, J. P.; van Haver, D.; Vandewalle, M.; Hulkenberg, A.; Veerman, W.; Nieuwenhuisen, R. *Tetrahedron Lett.* **1989**, *30*, 3873.
- 24. Bosmans, J. P.; van der Eycken, J.; Vandewalle, M.; Hulkenberg, A.; van Hes, R.; Veerman, W. *Tetrahedron Lett.* **1989**, *30*, 3877.
- 25. Hitotsuyanagi, Y.; Yamagami, K.; Fujii, A.; Naka, Y.; Ito, Y.; Tahana, T. *Bioorg. Med. Lett.* **1995**, *5*, 1039.
- 26. Hitotsuyanagi, Y.; Kobayashi, M.; Morita, H.; Itokawa, H.; Takeya, K. *Tetrahedron Lett.* **1999**, *40*, 9107.
- 27. Comins, D. L.; Hong, H.; Saha, J. K.; Jianhua, G. J. Org. Chem. 1994, 59, 5120.

- 28. Posner, G. H.; Canella, K. A. J. Am. Chem. Soc. 1985, 107, 2571.
- 29. Couture, A.; Deniau, E.; Grandclaudon, P.; Woisel, P. *Tetrahedron* **1996**, *52*, 4433.
- 30. Lebrun, S.; Couture, A.; Deniau, E.; Grandclaudon, P. *Tetrahedron* **1999**, *55*, 2659.
- 31. Sonesson, C.; Larhed, M.; Nyqvist, C.; Hallberg, A. J. Org. Chem. 1996, 61, 4756.
- 32. Sonesson, C.; Wikstroem, H.; Smith, M. W.; Svensson, K.; Carlsson, A.; Waters, N. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 241.
- 33. Itoh, N.; Sugasawa, S. Tetrahedron 1959, 6, 16.
- 34. Kametani, T.; Fukumoto, K. Heterocycles 1975, 3, 311.
- 35. Overman, L. E.; Kakimoto, M.; Okazaki, M. E.; Meier, G. P. J. Am. Chem. Soc. 1983, 105, 6622.

- 36. Miller, R. C. J. Org. Chem. 1959, 24, 2013.
- 37. Jogia, M. K.; Vakamoce, V.; Weavers, R. T. Aust. J. Chem. 1985, 38, 1009.
- 38. Heiser, B., Broger, E. A.; Crameri, Y. Tetrahedron: Asymmetry 1991, 2, 51.
- 39. Bradley, R. J. Chem. Soc., 1928, 1555.
- 40. Loriot, M.; Robin, P.; Brown, E. Tetrahedron 1984, 40, 2529.
- 41. Brown, E.; Loriot, M.; Robin, J.-P. *Tetrahedron Lett.* **1982**, *23*, 949.
- 42. Musser, J. H.; Chakraborty, U.; Bailey, K.; Sciortino, S.; Whyzmuzis, C. J. Med. Chem. 1987, 30, 62.
- 43. Léonce, S.; Pérez, V.; Casabianca-Pignède, M. R.; Anslett, M.; Bisagni, E.; Atassi, G. *Invest. New Drugs* **1996**, *14*, 169.